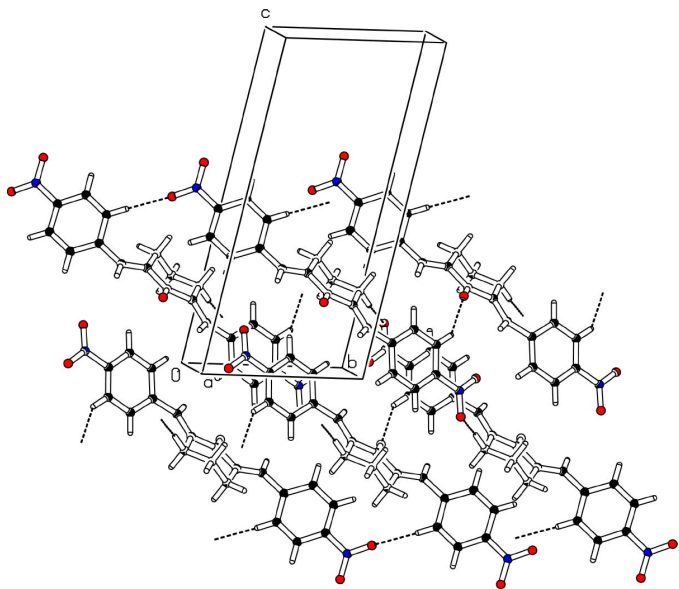


**Figure 1**

A general ORTEP-3 view (Farrugia, 1997) of (I), with displacement ellipsoids for non-H atoms drawn at the 50% probability level. For clarity, the H atoms are drawn as small spheres of arbitrary size.



**Figure 2**

A packing diagram (Spek, 2003) of (I). The weak C—H...O interactions are indicated by dashed lines. The interleaved stacking of nitroaryl groups can be seen intersecting the bottom C face of the unit cell.

respectively) causing the two aryl rings to twist out of the plane of the adjacent olefinic linkages and creating torsion angles for C6—C7—C8—C13 ( $\Theta_1$ ) and C2—C14—C15—C20 ( $\Theta_2$ ) of 38.5 (4) and  $-47.4$  (4) $^\circ$ , respectively. There are weak C—H...O interactions involving the carbonyl O atom and one O atom of each nitro group, as shown in Table 1. Furthermore, the relevant bond angles of (I) (Table 2) are all greater than  $120^\circ$ , confirming the repulsion between the aryl and cyclohexane rings.

In an attempt to ascertain the structural parameters contributing to the marked cytotoxicity of (I), its shape was compared with the less bioactive analogue (II). The  $\Theta_1$  and  $\Theta_2$  values of (II) are 40.1 (2) and  $-28.7$  (2) (Jia *et al.*, 1989), *i.e.* one value is very similar and the other is significantly larger for (I). In solution, the two angles should be of the same magnitude with opposite signs. The significant differences must be due to crystal packing effects. The nitroaryl group corresponding to  $\Theta_1$  is interleaved with equivalent nitroaryl groups in the crystal structure of (I) [interlayer distances of 3.376 (4) and 3.300 (4) Å], with some weak C—H...O bonds from C—H units to carbonyl and nitro O atoms (Fig. 2). The greater

lack of coplanarity of the aryl rings with the adjacent olefinic linkages in (I) than (II) may enable better alignments in narrow clefts at a binding site. Thus, in the future, the placement of nitro groups in the *ortho* and *meta* positions of the aryl rings, thereby creating even larger  $\Theta$  values, may lead to a correlation between the  $\Theta$  values and cytotoxic potencies. Second, as revealed in Table 2, certain bond angles are smaller in compound (I) than in (II). These data imply that the aryl rings are closer to the central alicyclic ring in (I) than in (II), which may be a contributing factor in the variation of potencies among the 2,6-bis(arylidene)cyclohexanones.

## Experimental

A solution of sodium hydroxide (0.01 mol) in water (1 ml) was added to a solution of cyclohexanone (0.01 mol) and 4-nitrobenzaldehyde (0.02 mol) in ethanol (20 ml) at room temperature. After stirring the reaction mixture at room temperature for 4 h, the precipitate was collected and recrystallized from acetone to give the title compound [m.p. 477 K; literature m.p. 473–476 K (Dibella, 1968)] in 65% yield.

### Crystal data

$C_{20}H_{16}N_2O_5$   
 $M_r = 364.35$   
 Triclinic,  $P\bar{1}$   
 $a = 7.5132$  (5) Å  
 $b = 7.8133$  (3) Å  
 $c = 16.1313$  (11) Å  
 $\alpha = 77.537$  (4) $^\circ$   
 $\beta = 89.546$  (3) $^\circ$   
 $\gamma = 65.600$  (3) $^\circ$   
 $V = 838.53$  (9) Å $^3$

$Z = 2$   
 $D_x = 1.443$  Mg m $^{-3}$   
 Mo  $K\alpha$  radiation  
 Cell parameters from 3465 reflections  
 $\theta = 1.0$ – $27.5^\circ$   
 $\mu = 0.11$  mm $^{-1}$   
 $T = 123$  (2) K  
 Chip, yellow  
 $0.20 \times 0.20 \times 0.10$  mm

### Data collection

Bruker–Nonius KappaCCD diffractometer  
 $\varphi$  scans and  $\omega$  scans with  $\kappa$  offsets  
 Absorption correction: none  
 6036 measured reflections  
 3290 independent reflections

2269 reflections with  $I > 2\sigma(I)$   
 $R_{int} = 0.040$   
 $\theta_{max} = 26.0^\circ$   
 $h = -9 \rightarrow 9$   
 $k = -9 \rightarrow 9$   
 $l = -19 \rightarrow 19$

### Refinement

Refinement on  $F^2$   
 $R[F^2 > 2\sigma(F^2)] = 0.056$   
 $wR(F^2) = 0.132$   
 $S = 1.09$   
 3290 reflections  
 244 parameters  
 H-atom parameters constrained

$w = 1/[\sigma^2(F_o^2) + (0.0405P)^2 + 0.5405P]$   
 where  $P = (F_o^2 + 2F_c^2)/3$   
 $(\Delta/\sigma)_{max} < 0.001$   
 $\Delta\rho_{max} = 0.22$  e Å $^{-3}$   
 $\Delta\rho_{min} = -0.23$  e Å $^{-3}$

**Table 1**

Hydrogen-bonding geometry (Å,  $^\circ$ ).

D—H...A	D—H	H...A	D...A	D—H...A
C5—H5A...O2 <sup>i</sup>	0.99	2.54	3.441 (3)	152
C12—H12...O1 <sup>ii</sup>	0.95	2.35	3.192 (3)	147
C20—H20...O4 <sup>iii</sup>	0.95	2.47	3.368 (3)	158

Symmetry codes: (i)  $-x, 3 - y, -z$ ; (ii)  $x - 1, 1 + y, z$ ; (iii)  $x, 1 + y, z$ .

**Table 2**  
Selected bond angles (°) for (I) and (II).

	(I)	(II) <sup>a</sup>
C3—C2—C14	124.2 (2)	125.5 (2)
C2—C14—C15	125.7 (2)	129.0 (2)
C14—C15—C20	120.5 (2)	123.0 (1)
C5—C6—C7	124.8 (2)	125.5 (1)
C6—C7—C8	127.5 (2)	130.5 (1)
C7—C8—C13	123.2 (2)	124.5 (2)

Note: (a) data taken from Jia *et al.* (1989).

H atoms were placed in calculated positions, with C—H distances ranging from 0.95 to 0.99 Å, and included in the refinement in riding-model approximation, with  $U_{\text{iso}}(\text{H})$  values constrained to be 1.2 times  $U_{\text{eq}}$  of the carrier atom for all H atoms.

Data collection: *COLLECT* (Nonius, 1998); cell refinement: *SCALEPACK* (Otwinowski & Minor, 1997); data reduction: *SCALEPACK* and *DENZO* (Otwinowski & Minor, 1997); program(s) used to solve structure: *SIR97* (Altomare *et al.*, 1999); program(s) used to refine structure: *SHELXL97* (Sheldrick, 1997); molecular graphics: *ORTEP-3 for Windows* (Farrugia, 1997) and *PLATON* (Spek, 2003); software used to prepare material for publication: *PLATON*.

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## References

- Altomare, A., Burla, M. C., Camalli, M., Cascarano, G., Giacovazzo, C., Guagliardi, A., Moliterni, A. G. G., Polidori, G. & Spagna, R. (1999). *J. Appl. Cryst.* **32**, 115–119.
- Dibella, E. P. (1968). US Patent No. 3 389 986, June 25, 1968; *Chem. Abstr.* (1968), **69**, 5182y.
- Dimmock, J. R., Kumar, P., Nazarali, A. J., Motaganahalli, N. L., Kowalchuk, T. P., Beazely, M. A., Quail, J. W., Oloo, E. O., Allen, T. M., Szydlowski, J., De Clercq, E. & Balzarini, J. (2000). *Eur. J. Med. Chem.* **35**, 967–977.
- Dimmock, J. R., Padmanilayam, M. P., Zello, G. A., Nienaber, K. H., Allen, T. M., Santos, C. L., De Clercq, E., Balzarini, J., Manavathu, E. K. & Stables, J. P. (2003). *Eur. J. Med. Chem.* **38**, 169–177.
- Dimmock, J. R., Zello, G. A., Oloo, E. O., Quail, J. W., Kraatz, H.-B., Perjési, P., Aradi, F., Takács-Novak, K., Allen, T. M., Santos, C. L., Balzarini, J., De Clercq, E. & Stables, J. P. (2002). *J. Med. Chem.* **45**, 3103–3111.
- Farrugia, L. J. (1997). *J. Appl. Cryst.* **30**, 565.
- Jia, A., Quail, J. W., Arora, V. K., & Dimmock, J. R. (1989). *Acta Cryst.* **C45**, 285–289.
- Nonius (1998). *COLLECT*. Nonius BV, Delft, The Netherlands.
- Otwinowski, Z. & Minor, W. (1997). *Methods in Enzymology*, Vol. 276, *Macromolecular Crystallography*, Part A, edited by C. W. Carter Jr and R. M. Sweet, pp. 307–326. New York: Academic Press.
- Quinn, F. R. & Milne, G. W. A. (1986). *Fundam. Appl. Toxicol.* **6**, 270–277.
- Sheldrick, G. M. (1997). *SHELXL97*. University of Göttingen, Germany.
- Spek, A. L. (2003). *J. Appl. Cryst.* **36**, 7–13.